

## ALKALINE BACILLUS AMYLASE

### CROSS-REFERENCE TO RELATED APPLICATIONS

This application is a continuation-in-part of application no. 09/249,558 filed on February 12, 1999 and claims priority under 35 U.S.C. 119 of Danish application no. 0228/98 filed on February 18, 1998, and U.S. provisional application no. 60/146,297, filed on February 27, 1998, the contents of which are fully incorporated by reference.

### FIELD OF INVENTION

The present invention relates to amylases having improved washing performance in an alkaline detergent solution at low temperature.

### BACKGROUND OF THE INVENTION

For a number of years  $\alpha$ -amylase enzymes have been used for a variety of different purposes, the most important of which are starch liquefaction, textile desizing, starch modification in the paper and pulp industry, and for brewing and baking. A further use of  $\alpha$ -amylases, which is becoming increasingly important is the removal of starchy stains during washing with a detergent at alkaline pH.

Examples of commercial  $\alpha$ -amylase products are Termamyl®, BAN® and Fungamyl®, all available from Novo Nordisk A/S, Denmark. These and similar products from other commercial sources have an acidic to a neutral pH optimum, typically in the range of from pH 5 to pH 7.5, and they do not display optimal activity in detergent solutions at alkaline pH.

WO 95/26397 discloses an  $\alpha$ -amylase from a *Bacillus* strain which has optimum activity at pH 8. WO 96/23873 describes variants of *Bacillus* amylases with improved performance under washing conditions.

US 5,147,796 describes an alkaline pullulanase having alpha-amylase activity. Fig. 2b of the document shows optimum amylase activity at pH 8-8.5.

M. Takagi et al., J. Ferment. Bioeng., vol 81, No. 6, 557-559 (1996) describe an alkaliphilic alpha-amylase-pullulanase from *Bacillus* sp. The enzyme has optimum amylase activity at pH 9, but the activity drops rapidly at higher pH, and the activity at pH 10 is lower than at pH 7.

It is an object of the present invention to provide novel  $\alpha$ -amylases with improved performance in alkaline solutions, especially in alkaline detergent solutions at pH around 9-11.

**SUMMARY OF THE INVENTION**

The present invention provides alpha-amylase which is:

- a) a polypeptide produced by *Bacillus* sp. NCIMB 40916, or
- b) a polypeptide having an amino acid sequence as shown in positions 1-556 of SEQ ID NO: 4, or
- c) a polypeptide encoded by the alpha-amylase encoding part of the DNA sequence cloned into a plasmid present in *Escherichia coli* DSM 13001, or
- d) an analogue of the polypeptide defined in (a) or (b) which:
  - i) is at least 60 % homologous with said polypeptide, or
  - ii) is derived from said polypeptide by substitution, deletion and/or insertion of one or several amino acids.

In an embodiment of the invention the alpha-amylase is the truncated mature alpha-amylase shown in SEQ ID NO: 2 (i.e., amino acids 1-517) encoded by nucleotides 94-1646 of the DNA sequence shown in SEQ ID NO: 1 and deposited as *Escherichia coli* DSM 12662.

In another aspect, the invention provides an  $\alpha$ -amylase having one or more of the following characteristics:

- an activity at pH 10.5 which is at least two times higher than the activity at pH 7.3 when measured at 37°C.

- an activity at pH 9.5 which is at least 4 times higher than the activity at pH 7.3 when measured at 37°C.

- an optimum pH of about 9.5 when measured at 37°C.

- a thermostability such that it retains more than 90 % of its activity after 20 minutes incubation at 25°C in a solution of 3 g/l of a test detergent containing 20% STPP, 25%  $\text{Na}_2\text{SO}_4$ , 15%  $\text{Na}_2\text{CO}_3$ , 20% LAS, 5% C12-C15 alcohol ethoxylate, 5%  $\text{Na}_2\text{SiO}_5$ , 0.3% NaCl at pH 10.5 and 6 degrees German hardness, and retains less than 90 % of its activity after 20 minutes incubation at 30°C in the same solution.

- a molecular weight of about 55 kDa as determined by SDS-PAGE.

- an iso-electric point of about 5 as determined by isoelectric focusing.

The invention also provides an isolated DNA sequence which encodes an alpha-amylase, wherein the alpha-amylase is that described above, or wherein the DNA sequence comprises:

- a) the DNA sequence shown in positions 94-1764 of SEQ ID NO: 3, or
- b) an analogue of the DNA sequence defined in a) which
  - i) is at least 60 % homologous with said DNA sequence, or
  - ii) hybridizes with said DNA sequence at least 55°C.

Other aspects of the invention provide a recombinant expression vector comprising the DNA sequence, and a cell transformed with the DNA sequence or the recombinant expression vector.

5 The invention also provides a method for producing an alpha-amylase by cultivating the cell and a detergent composition comprising said alpha-amylase.

### BRIEF DESCRIPTION OF DRAWINGS

The present invention is further illustrated with reference to the accompanying drawings, in which:

10 Fig. 1 shows a pH activity profile of the amylase from NCIMB 40916 compared to two prior-art amylases (SP722 and Termamyl).

Fig. 2 shows a temperature activity profile of the amylase from NCIMB 40916.

Fig. 3 shows the stability of the amylase from NCIMB 40916 after incubation at various temperatures.

15 Fig. 4 shows the cloning vector pSJ1678 described in Example 1.

Figs. 5 and 6 show results of washing tests described in Example 4.

### DETAILED DESCRIPTION OF THE INVENTION

#### Microbial source

20 The alpha-amylase of the invention may be derived from a strain of *Bacillus*. A preferred strain is *Bacillus* sp. NCIMB 40916. This strain was deposited on 28 January 1998 by the inventors under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure at the National Collections of Industrial and Marine Bacteria Limited (NCIMB), 23 St. Machar Drive, Aberdeen AB2 1RY, Scotland, United  
25 Kingdom. An *Escherichia coli* strain termed JA388 containing the truncated alpha amylase gene shown in SEQ ID NO: 1 cloned in plasmid pJA388 has also been deposited on 3 February 1999 under the terms of the Budapest Treaty with the Deutsche Sammlung von Microorganismen und Zellkulturen GmbH (DSMZ), Mascheroder Weg 1b, D-38124 Braunschweig DE, and given the accession  
30 number DSM 12662. Further, an *Escherichia coli* strain termed NN049489 containing the full length alpha amylase gene shown in SEQ ID NO: 3 cloned into plasmid pJA386 has also been deposited on 17 August, 1999, under the terms of the Budapest Treaty with the Deutsche Sammlung von Microorganismen und Zellkulturen GmbH (DSMZ), Mascheroder Weg 1b, D-38124 Braunschweig DE,  
35 and given the accession number DSM 13001. It is believed that SEQ ID NO: 3 and the deposited sequence are identical.

### Production of alpha-amylase

The alpha-amylase of the invention can be produced by cultivating a suitable amylase-producing strain of *Bacillus* or the transformed host cell of the invention in a suitable nutrient medium, and recovering the alpha-amylase from the culture medium.

The medium used to cultivate the cells may be any conventional medium suitable for growing the host cell in question and obtaining expression of the  $\alpha$ -amylase of the invention. Suitable media are available from commercial suppliers or may be prepared according to published recipes (e.g. as described in catalogues of the American Type Culture Collection).

The  $\alpha$ -amylase secreted from the host cells may conveniently be recovered from the culture medium by well-known procedures, including separating the cells from the medium by centrifugation or filtration, and precipitating proteinaceous components of the medium by means of a salt such as ammonium sulphate, followed by the use of chromatographic procedures such as ion exchange chromatography, affinity chromatography, or the like.

### Properties of alpha-amylase

A preferred alpha-amylase is derived from *Bacillus* sp. NCIMB 40916. It can be produced as described in the examples. The full length amino acid sequence of the amylase and the DNA encoding it are shown in SEQ ID NO: 3 and 4. The DNA and amino acid sequence of the truncated alpha-amylase is shown in SEQ ID NO: 1 and 3, respectively. The following characteristics were found for the full length amylase of the invention (purified alpha-amylase from NCIMB 40916):

A molecular weight of approximately 55 kDa as determined by SDS-PAGE using a Novex, 4-25% gradient gel.

A pI of approximately 5 was determined by isoelectric focusing (Ampholine PAG, pH 3.5-9.5).

A pH-activity curve is shown in Fig. 1, taking the activity at pH 7.3 as 100%. It was determined using the Phadebas assay using 50 mM Britten-Robinson buffer adjusted to various pH-values. For reference, the pH profiles of two prior-art *Bacillus* amylases (Termamyl derived from *B. licheniformis* and SP722 produced according to WO 95/26397) were measured under the same conditions are also shown in Figure 1. Figure 1 shows that the amylase of the invention has about 10 times higher activity than the two prior-art amylases at pH 9.5. Fig. 1 also shows that the optimum activity is about pH 9.5.

A temperature-activity curve was measured using the Phadebas assay at various temperatures with 50 mM Britten-Robinson buffer adjusted to pH 9.5. The

results are shown in Fig. 2. It is seen from Fig. 2 that the amylase of the invention has optimum activity at about 55°C.

Stability of the amylase was measured at pH 10.5 (50 mM CAPS) at 22, 37, 45, 55 and 65 °C after 30 minutes incubation. The enzyme was diluted to 40 NU/ml in buffer and 25 microlitre sample was incubated at the respective temperatures. After 30 minutes, the samples was chilled on ice and residual activity was determined. A stability profile of the amylase of the invention and a prior-art amylase (SP722) is shown in Figure 3.

Stability was tested by incubating 40 NU/ml of amylase in a solution of 3 g/l of the A/P model detergent described above, at pH 10.5 and 6°dH (German hardness, Ca:Mg 2:1). After the incubation, the residual activity was measured by Phadebas at pH 7.3. The results were 95 % residual activity after incubation at 25°C, and 87 % at 30°C.

### Homology of polypeptide and DNA sequence

The amino acid sequence homology may be determined as the degree of identity between the two sequences indicating a derivation of the first sequence from the second. The homology may suitably be determined by means of computer programs known in the art. Thus, FASTA provided in GCG version 8 (Needleman, S.B. and Wunsch, C.D., (1970), Journal of Molecular Biology, 48, 443-453) may be used with the following settings: Scoring matrix: GenRunData:blosum50.cmp, Variable pamfactor used Gap creation penalty: 12, Gap extension penalty: 2. Alternatively, Gap from GCG version 9 may be used with a translated version 8 peptide scoring matrix, a gap creation penalty of 30, a gap extension penalty of 1 using ntol's matrix (<http://plasmid/~bioweb/matrix/>) without end gap penalty.

The amino acid sequence exhibits a degree of identity preferably of at least 60%, preferably at least 70%, more preferably at least 80%, especially at least 90% or at least 95%, with the amino acid sequence shown in positions 1-556 of SEQ ID NO: 4.

The DNA sequence homology may be determined as the degree of identity between the two sequences indicating a derivation of the first sequence from the second. The homology may suitably be determined by means of computer programs known in the art such as GAP provided in the GCG program package (described above). Thus, Gap GCGv8 may be used with the following default parameters: GAP creation penalty of 5.0 and GAP extension penalty of 0.3, default scoring matrix . GAP uses the method of Needleman/Wunsch/Sellers to make alignments.

The DNA construct of the present invention comprises a DNA sequence exhibiting a degree of identity preferably of at least 60%, preferably at least 70%, more preferably at least 80%, especially at least 90% or at least 95%, with the nucleic acid sequence shown in positions 94-1764 of SEQ ID NO: 3.

## 5 Hybridization

The hybridization is used to indicate that a given DNA sequence is analogous to a nucleotide probe corresponding to SEQ ID NO: 3. The hybridization conditions are described in detail below.

Suitable conditions for determining hybridization between a nucleotide probe and a homologous DNA or RNA sequence involves presoaking of the filter containing the DNA fragments or RNA to hybridize in 5 x SSC (standard saline citrate) for 10 min, and prehybridization of the filter in a solution of 5 x SSC (Sambrook et al. 1989), 5 x Denhardt's solution (Sambrook et al. 1989), 0.5 % SDS and 100 µg/ml of denatured sonicated salmon sperm DNA (Sambrook et al. 1989), followed by hybridization in the same solution containing a random-primed (Feinberg, A. P. and Vogelstein, B. (1983) *Anal. Biochem.* 132:6-13), <sup>32</sup>P-dCTP-labeled (specific activity > 1 x 10<sup>9</sup> cpm/µg ) probe for 12 hours at approx. 45°C. The filter is then washed two times for 30 minutes in 2 x SSC, 0.5 % SDS at a temperature of at least 55°C, more preferably at least 60°C, more preferably at least 65°C, even more preferably at least 70°C, especially at least 75°C.

Molecules to which the oligonucleotide probe hybridizes under these conditions are detected using a x-ray film.

## Recombinant expression vector

The expression vector of the invention typically includes control sequences encoding a promoter, operator, ribosome binding site, translation initiation signal, and, optionally, a repressor gene or various activator genes.

The recombinant expression vector carrying the DNA sequence encoding the α-amylase of the invention may be any vector which may conveniently be subjected to recombinant DNA procedures, and the choice of vector will often depend on the host cell into which it is to be introduced. Thus, the vector may be an autonomously replicating vector, i.e., a vector which exists as an extrachromosomal entity, the replication of which is independent of chromosomal replication, e.g., a plasmid, a bacteriophage or an extrachromosomal element, minichromosome or an artificial chromosome. Alternatively, the vector may be one which, when introduced into a host cell, is integrated into the host cell genome and replicated together with the chromosome(s) into which it has been integrated.

In the vector, the DNA sequence should be operably connected to a suitable promoter sequence. The promoter may be any DNA sequence which shows transcriptional activity in the host cell of choice and may be derived from genes encoding proteins either homologous or heterologous to the host cell.

5 Examples of suitable promoters for directing the transcription of the DNA sequence encoding an  $\alpha$ -amylase of the invention, especially in a bacterial host, are the promoter of the *lac* operon of *E.coli*, the *Streptomyces coelicolor* agarase gene *dagA* promoters, the promoters of the *Bacillus licheniformis*  $\alpha$ -amylase gene (*amyL*), the promoters of the *Bacillus stearothermophilus* maltogenic amylase

10 gene (*amyM*), the promoters of the *Bacillus Amyloliquefaciens*  $\alpha$ -amylase (*amyQ*), the promoters of the *Bacillus subtilis* xylA and xylB genes etc. For transcription in a fungal host, examples of useful promoters are those derived from the gene encoding *A. oryzae* TAKA amylase, *Rhizomucor miehei* aspartic proteinase, *A. niger* neutral  $\alpha$ -amylase, *A. niger* acid stable  $\alpha$ -amylase, *A. niger* glucoamylase,

15 *Rhizomucor miehei* lipase, *A. oryzae* alkaline protease, *A. oryzae* triose phosphate isomerase or *A. nidulans* acetamidase.

The expression vector of the invention may also comprise a suitable transcription terminator and, in eukaryotes, polyadenylation sequences operably connected to the DNA sequence encoding the  $\alpha$ -amylase of the invention. Termination and polyadenylation sequences may suitably be derived from the same

20 sources as the promoter.

The vector may further comprise a DNA sequence enabling the vector to replicate in the host cell in question. Examples of such sequences are the origins of replication of plasmids pUC19, pACYC177, pUB110, pE194, pAMB1 and

25 pIJ702.

The vector may also comprise a selectable marker, e.g. a gene the product of which complements a defect in the host cell, such as the *dal* genes from *B. subtilis* or *B. licheniformis*, or one which confers antibiotic resistance such as ampicillin, kanamycin, chloramphenicol or tetracyclin resistance. Furthermore, the

30 vector may comprise *Aspergillus* selection markers such as *amdS*, *argB*, *niaD* and *sC*, a marker giving rise to hygromycin resistance, or the selection may be accomplished by co-transformation, e.g., as described in WO 91/17243.

While intracellular expression may be advantageous in some respects, e.g. when using certain bacteria as host cells, it is generally preferred that the

35 expression is extracellular.

Procedures suitable for constructing vectors of the invention encoding an  $\alpha$ -amylase, and containing the promoter, terminator and other elements, respectively, are well known to persons skilled in the art [cf., for instance, Sambrook et al. (1989)].

## Host cells

The cell of the invention, either comprising a DNA construct or an expression vector of the invention as defined above, is advantageously used as a host cell in the recombinant production of the  $\alpha$ -amylase of the invention. The cell  
5 may be transformed with the DNA construct of the invention encoding the amylase, conveniently by integrating the DNA construct (in one or more copies) in the host chromosome. This integration is generally considered to be an advantage as the DNA sequence is more likely to be stably maintained in the cell. Integration of the DNA constructs into the host chromosome may be performed according to conven-  
10 tional methods, e.g. by homologous or heterologous recombination. Alternatively, the cell may be transformed with an expression vector as described above in connection with the different types of host cells.

The cell of the invention may be a cell of a higher organism such as a mammal or an insect, but is preferably a microbial cell, e.g. a bacterial or a fungal  
15 (including yeast) cell.

Examples of bacterial host cells which, on cultivation, are capable of producing the enzyme of the invention are gram-positive bacteria such as strains of *Bacillus*, such as strains of *B. subtilis*, *B. licheniformis*, *B. lentus*, *B. clausii*, *B. brevis*, *B. stearothermophilus*, *B. alkalophilus*, *B. amyloliquefaciens*, *B. coagulans*,  
20 *B. circulans*, *B. lautus*, *B. megaterium* or *B. thuringiensis*, or strains of *Streptomyces*, such as *S. lividans* or *S. murinus*, or gram-negative bacteria such as *Escherichia coli*. The transformation of the bacteria may be effected by protoplast transformation, electroporation, conjugation, or by using competent cells in a manner known per se (cf. Sambrook et al., supra).

The yeast organism may favourably be selected from a species of *Saccharomyces* or *Schizosaccharomyces*, e.g. *Saccharomyces cerevisiae*. The filamentous fungus may advantageously belong to a species of *Aspergillus*, e.g. *Aspergillus oryzae* or *Aspergillus niger*. Fungal cells may be transformed by a process involving protoplast formation and transformation of the protoplasts followed  
30 by regeneration of the cell wall in a manner known *per se*. A suitable procedure for transformation of *Aspergillus* host cells is described in EP 238 023.

## Industrial Applications

Owing to their activity at alkaline pH values, the  $\alpha$ -amylases of the invention are well suited for use in a variety of industrial processes, in particular the enzyme  
35 finds potential applications as a component in detergents, e.g. laundry and hard surface cleaning detergent compositions, but it may also be useful in the production of sweeteners and ethanol from starch. Thus, it may be used in conventional starch-



converting processes, such as liquefaction and saccharification processes described in US Patent No. 3,912,590 and EP patent publications Nos. 252,730 and 63,909.

The alkaline  $\alpha$ -amylase of the invention may also be used in the production of lignocellulosic materials, such as pulp, paper and cardboard, from starch reinforced waste paper and cardboard, especially where repulping occurs at pH above 7 and where amylases can facilitate the disintegration of the waste material through degradation of the reinforcing starch. The  $\alpha$ -amylase of the invention is especially useful in a process for producing a papermaking pulp from starch-coated printed paper. The process may be performed as described in WO 95/14807, comprising the following steps:

- a) disintegrating the paper to produce a pulp,
- b) treating with a starch-degrading enzyme before, during or after step a), and
- c) separating ink particles from the pulp after steps a) and b).

The  $\alpha$ -amylases of the invention may also be very useful in modifying starch where enzymatically modified starch is used in papermaking together with alkaline fillers such as calcium carbonate, kaolin and clays. With the alkaline  $\alpha$ -amylases of the invention it becomes possible to modify the starch in the presence of the filler thus allowing for a simpler integrated process.

The  $\alpha$ -amylase of the invention may also be very useful in textile desizing. In the textile processing industry,  $\alpha$ -amylases are traditionally used as auxiliaries in the desizing process to facilitate the removal of starch-containing size which has served as a protective coating on weft yarns during weaving. Complete removal of the size coating after weaving is important to ensure optimum results in the subsequent processes, in which the fabric is scoured, bleached and dyed. Enzymatic starch break-down is preferred because it does not involve any harmful effect on the fiber material. In order to reduce processing cost and increase mill throughput, the desizing processing is sometimes combined with the scouring and bleaching steps. In such cases, non-enzymatic auxiliaries such as alkali or oxidation agents are typically used to break down the starch, because traditional  $\alpha$ -amylases are not very compatible with high pH levels and bleaching agents. The non-enzymatic breakdown of the starch size does lead to some fiber damage because of the rather aggressive chemicals used. Accordingly, it would be desirable to use the  $\alpha$ -amylases of the invention as they have an improved performance in alkaline solutions. The  $\alpha$ -amylases may be used alone or in combination with a cellulase when desizing cellulose-containing fabric or textile.

The  $\alpha$ -amylases of the invention may also be very useful in a beer-making process; the  $\alpha$ -amylases will typically be added during the mashing process.

## Detergent Compositions

According to the invention, the  $\alpha$ -amylase may typically be a component of a detergent composition, e.g., a laundry detergent composition. As such, it may be included in the detergent composition in the form of a non-dusting granulate, a stabilized liquid, or a protected enzyme. Non-dusting granulates may be produced, e.g., as disclosed in US 4,106,991 and 4,661,452 (both to Novo Industri A/S) and may optionally be coated by methods known in the art. Examples of waxy coating materials are poly(ethylene oxide) products (polyethyleneglycol, PEG) with mean molecular weights of 1000 to 20000; ethoxylated nonylphenols having from 16 to 50 ethylene oxide units; ethoxylated fatty alcohols in which the alcohol contains from 12 to 20 carbon atoms and in which there are 15 to 80 ethylene oxide units; fatty alcohols; fatty acids; and mono- and di- and triglycerides of fatty acids. Examples of film-forming coating materials suitable for application by fluid bed techniques are given in patent GB 1483591. Liquid enzyme preparations may, for instance, be stabilized by adding a polyol such as propylene glycol, a sugar or sugar alcohol, lactic acid or boric acid according to established methods. Other enzyme stabilizers are well known in the art. Protected enzymes may be prepared according to the method disclosed in EP 238,216.

The properties of the alpha-amylase of the invention make it particularly suitable for use in alkaline detergents, e.g. at pH 9.5-10.5, and for washing at low temperatures, e.g. 20-40°C.

The detergent composition of the invention may be in any convenient form, e.g. as powder, granules, paste or liquid. A liquid detergent may be aqueous, typically containing up to 70% water and 0-30% organic solvent, or non-aqueous.

The detergent composition comprises one or more surfactants, each of which may be anionic, nonionic, cationic, or amphoteric (zwitterionic). The detergent will usually contain 0-50% of anionic surfactant such as linear alkylbenzenesulfonate (LAS), alpha-olefinsulfonate (AOS), alkyl sulfate (fatty alcohol sulfate) (AS), alcohol ethoxysulfate (AEOS or AES), secondary alkanesulfonates (SAS), alpha-sulfo fatty acid methyl esters, alkyl- or alkenylsuccinic acid, or soap. It may also contain 0-40% of nonionic surfactant such as alcohol ethoxylate (AEO or AE), alcohol propoxylate, carboxylated alcohol ethoxylates, nonylphenol ethoxylate, alkylpolyglycoside, alkyltrimethylamine oxide, ethoxylated fatty acid monoethanolamide, fatty acid monoethanolamide, or polyhydroxy alkyl fatty acid amide (e.g. as described in WO 92/06154).

The detergent composition may additionally comprise one or more other enzymes, such as pullulanase, esterase, lipase, cutinase, protease, cellulase, peroxidase, or oxidase, e.g., laccase.

Normally the detergent contains 1-65% of a detergent builder or complexing agent such as zeolite, diphosphate, triphosphate, phosphonate, citrate, nitrilotriacetic acid (NTA), ethylenediaminetetraacetic acid (EDTA), diethylenetriaminepentaacetic acid (DTMPA), alkyl- or alkenylsuccinic acid, soluble silicates or  
5 layered silicates (e.g. SKS-6 from Hoechst).

The detergent builders may be subdivided into phosphorus-containing and non-phosphorous-containing types. Examples of phosphorus-containing inorganic alkaline detergent builders include the water-soluble salts, especially alkali metal pyrophosphates, orthophosphates, polyphosphates and phosphonates. Examples  
10 of non-phosphorus-containing inorganic builders include water-soluble alkali metal carbonates, borates and silicates as well as layered disilicates and the various types of water-insoluble crystalline or amorphous aluminosilicates of which zeolites is the best known representative.

Examples of suitable organic builders include alkali metal, ammonium or  
15 substituted ammonium salts of succinates, malonates, fatty acid malonates, fatty acid sulphonates, carboxymethoxy succinates, polyacetates, carboxylates, polycarboxylates, aminopolycarboxylates and polyacetyl carboxylates. The detergent may also be unbuilt, i.e. essentially free of detergent builder.

The detergent may comprise one or more polymers. Examples are  
20 carboxymethylcellulose (CMC), poly(vinylpyrrolidone) (PVP), polyethyleneglycol (PEG), poly(vinyl alcohol) (PVA), polycarboxylates such as polyacrylates, polymaleates, maleic/acrylic acid copolymers and lauryl methacrylate/acrylic acid copolymers.

The detergent composition may contain bleaching agents of the  
25 chlorine/bromine-type or the oxygen-type. The bleaching agents may be coated or encapsulated.

Examples of inorganic chlorine/bromine-type bleaches are lithium, sodium or calcium hypochlorite or hypobromite as well as chlorinated trisodium phosphate. Examples of organic chlorine/bromine-type bleaches are heterocyclic N-bromo and  
30 N-chloro imides such as trichloroisocyanuric, tribromoisocyanuric, dibromoisocyanuric and dichloroisocyanuric acids, and salts thereof with water solubilizing cations such as potassium and sodium. Hydantoin compounds are also suitable. The bleaching system may also comprise peroxyacids of, e.g., the amide, imide, or sulfone type.

The oxygen-type bleach may be an inorganic persalt, preferably with a  
35 bleach activator, or a peroxy acid compound. Examples of inorganic persalts are alkali metal perborates, both tetrahydrates and monohydrates, alkali metal percarbonates, persilicates and perphosphates. The activator may be tetraacetythylenediamine (TAED) or nonanoyloxybenzenesulfonate (NOBS).

The enzymes of the detergent composition of the invention may be stabilized using conventional stabilizing agents, e.g. a polyol such as propylene glycol or glycerol, a sugar or sugar alcohol, lactic acid, boric acid, or a boric acid derivative such as, e.g., an aromatic borate ester, and the composition may be formulated as described in, e.g., WO 92/19709 and WO 92/19708. The enzymes of the invention may also be stabilized by adding reversible enzyme inhibitors, e.g., of the protein type as described in EP 0 544 777 B1.

The detergent may also contain other conventional detergent ingredients such as, e.g., fabric conditioners including clays, deflocculant material, foam boosters, foam depressors, suds suppressors, anti-corrosion agents, soil-suspending agents, anti-soil-redeposition agents, dyes, dehydrating agents, bactericides, optical brighteners, or perfume.

The pH (measured in aqueous solution at use concentration) will usually be neutral or alkaline, e.g. in the range of 7-11.

More specifically, the alpha-amylase of the invention may be incorporated in any of the detergent formulations described in WO 96/23873.

The  $\alpha$ -amylases of the invention may be incorporated in concentrations conventionally employed in detergents. It is at present contemplated that, in the detergent composition of the invention, the  $\alpha$ -amylase may be added in an amount corresponding to 0.00001-1 mg (calculated as pure enzyme protein) of  $\alpha$ -amylase per liter of wash liquor.

The present invention is further illustrated in the following examples which are not intended to be in any way limiting to the scope of the invention as claimed.

## EXAMPLES

### Host organism:

*Escherichia coli* SJ2 is described in Diderichsen, B., Wedsted, U., Hedegaard, L., Jensen, B.R., Sjøholm, C. (1990) Journal of Bacteriology, Vol.172, No. 8, p. 4315-4321.

### Plasmids:

The gene bank vector was pSJ1678 which is further disclosed in WO94/19454 which is hereby incorporated by reference.

The gene bank vector pSJ1678 is further disclosed in WO 94/19454 which is hereby incorporated by reference.

### Model detergent:

A/P (Asia/Pacific) Model Detergent has the following composition: 20% STPP (sodium tripolyphosphate), 25% Na<sub>2</sub>SO<sub>4</sub>, 15% Na<sub>2</sub>CO<sub>3</sub>, 20% LAS (linear

alkylbenzene sulfonate, Nansa 80S), 5% C<sub>12</sub>-C<sub>15</sub> alcohol ethoxylate (Dobanol 25-7), 5% Na<sub>2</sub>Si<sub>2</sub>O<sub>5</sub>, 0.3% NaCl.

### Example 1

#### Cloning of alpha-amylase into *E. coli*

5 DNA was isolated from *Bacillus* sp. NCIMB 40916 by the method of Pitcher, D. G., Saunders, N. A., and Owen, R. J. (1989) Lett. Appl. Microbiol. 8, 151-156. Chromosomal DNA was partially digested with the restriction enzyme Sau3AI. The fragments were cloned into the BamHI site of the cloning vector pSJ1678, as shown in Figure 4 and transformed into *Escherichia coli* SJ2, thereby  
10 creating a gene library of *Bacillus* sp. NCIMB 40916.

This gene library was screened for alpha amylase activity and the strain showing alpha amylase activity was termed *Escherichia coli* strain NN049489 comprises the full length alpha-amylase of the invention. This strain was deposited and given the accession number DSM 13001. *Escherichia coli* strain DSM 13001  
15 (NN049489) harbouring the plasmid termed pJA386 encoding the full length alpha amylase was used for DNA sequencing. Further, *Escherichia coli* strain JA388 comprises a truncated alpha-amylase. This was deposited and given the accession number DSM 12662. *Escherichia coli* strain JA388 harbours the plasmid termed pJA388 encoding the truncated alpha amylase.

### 20 Example 2

#### Sequencing of DNA and alpha-amylase

The full length alpha amylase gene cloned in plasmid pJA386 was characterized by DNA sequencing by primer walking, using the Taq deoxyterminal cycle sequencing kit (Perkin Elmer, USA), fluorescent labeled terminators and  
25 appropriate oligonucleotides as primers.

Analysis of the sequence data was performed according to Devereux et al. (1984) Nucleic Acids Res. 12, 387-395. The sequence corresponds to the DNA sequence shown in SEQ ID NO: 3. The predicted protein sequence of the full length alpha amylase including the signal peptide and the mature alpha amylase  
30 are presented in SEQ ID NO: 4. The deduced N-terminal sequence was verified by sequencing 34 amino acids at the N-terminal of the protein.

### Example 3

#### Production of alpha-amylase

*E. coli* DSm13001 (NN049489) harboring plasmid pJA386 was cultivated  
35 over night in LB-broth containing chloramphenicol 10 µg/ml, 37 °C, 250 rpm. Cells were harvested from 2.7 l culture broth by centrifugation at 6000 rpm for 15

minutes. The intracellular located amylase was released from the cells by using the following osmotic shock procedure:

1) Cells were resuspended and washed in 500 ml 10 mM Tris- HCl, pH 7.0 (EKV-buffer) followed by centrifugation.

5        2) Cells were resuspended in 75 ml 20% sucrose, 30 mM Tris-HCl pH 8, 1 mM EDTA and added Lysozyme to a concentration of 5 mg/ml.

3) The solution was incubated 15 min on ice followed by centrifugation. The majority of the amylase was now contained in the supernatant.

10        The supernatant was dialyzed overnight against 10 l in EKV-buffer with one buffer change to decrease the high concentration of sucrose. The solution was filtered through a 0.45 µm membrane vacuum filter.

15        The enzyme solution was applied on a Pharmacia Q Sepharose column FF previously equilibrated in EKV-buffer, pH 7, and the column was washed with EKV-buffer. Bound proteins were eluted with a linear NaCl gradient from 0-500 mM over 10 column volumes. Amylase containing fractions were pooled and dialyzed against EKV-buffer over night.

20        The solution was then applied on a Q Sepharose column FF previously equilibrated in EKV-buffer, pH 8, the column was washed with EKV-buffer, and the amylase was eluted with a linear NaCl gradient from 0-500 mM over 10 column volumes. Amylase containing fractions were pooled.

25        A Phenyl Superose column previously equilibrated in EKV-buffer, pH 8 containing 1 M ammonium sulfate was loaded with the enzyme solution added ammonium sulfate to a concentration of 1 M. Unbound material was washed out with the ammonium sulfate buffer and the column was eluted with a linear NaCl gradient from 1-0 M ammonium sulfate over 20 column volumes. Amylase containing fractions were pooled.

The purified amylase was analyzed by SDS-PAGE and only one band was obtained after staining with Coomassie Blue.

#### **Example 4**

##### **30    Washing test of the full length alpha-amylase (SEQ ID NO: 4)**

Washing performance was evaluated by washing soiled test swatches for 15 minutes at 25°C in a detergent solution with the alpha-amylase of the invention.

35        The detergent used was the A/P Model Detergent described above at 3 g/l having pH 10.5, or a commercial detergent from Malaysia (FAB Total from Colgate) at 3 g/l having a pH of approx. 9.7. The purified amylase of Example 3 was added to the detergent solution at the concentration indicated below. The test swatches were soiled with orange rice starch (CS-28 swatches available from CFT, Center for Test Material, Holland).

After washing, the swatches were evaluated by measuring the remission at 460 nm. The results are expressed as  $\Delta R$  = remission of the swatch washed with the alpha-amylase minus the remission of a swatch washed at the same conditions without the alpha-amylase.

Alpha-amylase concentration (mg enzyme protein / l)	$\Delta R$ Model detergent	$\Delta R$ Malaysian detergent
0 (reference)	= 0	= 0
0.1	0.9	1
0.2	1.9	1.3
0.5	3	2.4
1.5	4.1	4.3

- 5 The results are shown in Figures 5 and 6. The results demonstrate that the alpha-amylase of the invention is effective in both detergents at highly alkaline pH.